SOP: **Propagation of human iris pigment epithelial cells (HIPEpiC, ScienCell)**
Date modified: **12/11/09**
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Ordering Information

Human iris pigment epithelial cells (HIPEpiC) may be ordered as frozen ampoules. Each vial contains >5 x 10^5 cells in a 1ml volume.

To order frozen ampoules + media:

Name:	HIPEpiC – Human Iris Pigment Epithelial Cells
Item #:	6560 (HIPEpiC - Cryopreserved ampoule)
	4101 (EpiCM, Epithelial Cell Medium)

Notes:

The number of media kits purchased depends on the target number of cells to be generated. A rule of thumb is 10 media kits for every initial cryopreserved ampoule. It is strongly recommended to purchase all of the media that will be required for a complete expansion series, since media supply may be erratic. All culture flasks must be coated with poly-L-lysine (2ug/cm^2).

Materials List

1. EpiCM Epithelial Cell Medium (ScienCell, Cat# 4101)
2. Poly-L-Lysine (10mg/ml, ScienCell, Cat# 0413)
3. Phosphate Buffered Saline (1X PBS) (prepared from 10X stock Cellgro, Cat# 46-013-CM by dilution with sterile deionized water)
4. Accutase Enzyme Cell Detachment Medium (EBioscience, Cat# 00-4555)
5. T225 culture flasks
6. Graduated pipets (1, 5, 10, 25, 50mL)
7. Pen-Strep solution (if required; ScienCell typically supplies antibiotics)
8. Hemocytometer
9. Micropipet w/ P20 tips
10. Microscope

Procedure

A. Initiation of culture from cryopreserved cells

1) Rapidly thaw cells by holding vial and gently rotating in 37°C water bath.
2) As soon as ice crystals disappear, dispense contents of vial into a flask at 5,000 cells/cm^2 density.
3) Let cells recover for 16 hours in a 37°C, 5% CO_2 humidified incubator.
4) The next morning, the diluted DMSO-containing shipping/cryopreservation medium is aspirated from the cell layer and replaced with fresh medium.

B. Sub-culture

1) Propagate cells until density reaches 90% confluence.
2) Aspirate medium.
3) Wash cells with warm 1X PBS.
4) Add 15mLs of Accutase and return to incubator for 10-15 minutes, or until cells detach.
5) Immediately remove cells, rinse flask with warm 1X PBS to collect residual cells, and pellet at 500 x g for 5 minutes (4^oC).
6) Gently re-suspend cell pellet in warm medium.
7) Count cells with hemocytometer.
8) Add warmed medium to flasks.
9) Seed flasks at 5,000 cells/cm^2
10) Record each subculture event as a passage.

C. Maintenance

1) Change media the day after seeding and every OTHER day thereafter.
2) Increase media volume as confluency increases (volumes assume the use of
3) T225 flasks):
    a. 25 % = 1mL/5 cm2
    b. 25-45% = 1.5mL/ 5 cm2
    c. 45%+ = 2mL/ 5 cm2.
4) Per the above an exemplary schedule might be:
    a. day 1, plate into T225: use 50 mls of media.
    b. day 2, change media, use 50 mls of media
    c. day 4, change media, use 100 mls of media (if confluency is >50%)
    d. day 6, change media, use 100 mls of media (or harvest if ready)
    e. day 7 or 8 (harvest when cells reach 6 x 10^6 cells/flask

D. Harvest

1) Pass cells 3-4 times until the desired cell number is achieved (primary cells will senesce after 4-5 passages).
2) Remove cells from flasks according to protocol described above under 'Sub-culture'.
3) Examine viability using Trypan blue staining (SOP TP-7).