

SOP: Propagation of HL-60
Date modified: 4/20/2009
Modified by: J. Goldy/M. Dorschner

Ordering Information

HL-60 may be ordered from ATCC as a frozen ampoule.

Name: HL-60, acute promyelocytic leukemia
ATCC #: CCL-240

Notes:

This cell line grows in suspension.

Materials List

1. DMEM with 2mM L-glutamine (cellgro Cat# 10-013-CM)
2. Fetal Bovine Serum (cellgro Cat# 35-016-CV)
3. T75 & T225 culture flasks
4. Graduated pipets (1, 5, 25mL)
5. Penicillin-Streptomycin Solution (100X) (Cellgro Cat# 30-002-CI)
6. Hemocytometer
7. Micropipet w/ P20 tips
8. Microscope

Growth Media for HL-60

DMEM with 2mM L-glutamine (cellgro Cat# 10-013-CM)
20% FBS
1x Pen-Strep

Procedure

A. Receipt of Frozen cells and starting cell cultures.

- 1) Immediately place frozen cells in liquid nitrogen storage incubator.
- 2) Quickly thaw ampoule in 37°C water bath.
- 3) Transfer thawed cells to a T75 flask at 1×10^5 density in warm growth medium.
- 4) Allow cells to recover over night in 37°C, 5% CO₂ humidified incubator.
- 5) Pour off medium the next day, replace with fresh medium and return to incubator.

B. Sub-culture

- 1) Propagate cells until density reaches $\sim 8 \times 10^5$ (not to exceed 1×10^6).
- 2) Decant medium.
- 3) Wash cells with warm 1X PBS.
- 4) Immediately remove cells and pellet at 500 xg for 3 minutes (4°C)
- 5) Wash cells 2X with 1X PBS.
- 6) Gently re-suspend cell pellet in warm medium.
- 7) Seed at density of 1×10^5 .
- 8) Record each subculture event as a passage.

C. Maintenance

- 1) Change media the day after seeding and every 2-3 days thereafter.

D. Harvest

- 1) Do not use cells that have been passed more than 25 times
- 2) Remove cells from flasks according to protocol described above under 'sub-culturing'
- 3) Examine viability using trypan blue staining.