

**SOP:** Propagation of Human Mammary Epithelial Cells (HMEC, Lonza Biosciences)  
**Date modified:** 6/1/2010  
**Modified by:** T. Canfield (UW)

### **Ordering Information**

Human Mammary Epithelial Cells (HMEC) may be ordered either as frozen ampoules or as starter cultures. The former contain  $\sim 0.5-1 \times 10^6$  cells; the latter are initiated at Lonza and sent in a T225 flask containing  $6-7 \times 10^6$  cells.

To order frozen ampoules + media:

Name: HMEC – Human Mammary Epithelial Cells  
Item #: CC-2551 (HMEC in MEGM® - Cryopreserved ampule)  
CC-3150 (MEGM BulletKit = CC-3151 + CC-4136)

To order starter cultures:

Name: HMEC – Human Mammary Epithelial Cells  
Item #: CC2551T225 (HMEC in MEGM® T225 Flask)  
CC-3150 (MEGM BulletKit = CC-3151 + CC-4136)

### **Notes:**

The number of BulletKits purchased depends on the target number of cells to be generated. A rule of thumb is 10 BulletKits for every initial T225 flask of cells. It is strongly recommended to purchase all of the media that will be required for a complete expansion series, since media supply may be erratic.

### **Materials List**

1. Cell-type specific medium (BulletKits – Lonza Biosciences)
2. T225 culture flasks
3. Graduated pipets (1, 5, 10, 25, 50mL)
4. Pen-Strep solution (if required; Lonza typically supplies antibiotics)
5. Phosphate Buffered Saline (1X PBS) (Cellgro, Cat# 21-040-CM)
6. Accutase Enzyme Cell Detachment Medium (EBiosciences Cat# 00-4555)
7. Hemocytometer
8. Micropipet w/ P20 tips
9. Microscope

### **Procedure**

#### **A. Receipt of proliferating cells**

- 1) Swab down flask with 70% ethanol.
- 2) Equilibrate for 3-4 hours in 37°C, 5% CO<sub>2</sub> humidified incubator.
- 3) Remove shipping medium. Replace with fresh medium and return to incubator.

#### **B. Sub-culture**

- 1) Propagate cells until density reaches 60-80% confluence.

- 2) Aspirate medium.
- 3) Wash cells with warm 1X PBS.
- 4) Add 15mL of Accutase and return to incubator for 10-15 minutes, or until cells detach.
- 5) Immediately remove cells, rinse flask with warm 1X PBS to collect residual cells, and pellet at 500 x g for 5 minutes (4°C).
- 6) Gently re-suspend cell pellet in warm medium.
- 7) Count cells with hemocytometer.
- 8) Add warmed medium to flasks.
- 9) Seed flasks at **2,500 cells/cm<sup>2</sup> density**.
- 10) Record each subculture event as a passage.

### C. Maintenance

- 1) Change media the day after seeding and every OTHER day thereafter.
- 2) Increase media volume as confluency increases (volumes assume the use of
- 3) T225 flasks):
  - a. 25% = 1mL/5 cm<sup>2</sup>
  - b. 25-45% = 1.5mL/5 cm<sup>2</sup>
  - c. 45%+ = 2mL/5 cm<sup>2</sup>.
- 4) Per the above an exemplary schedule might be:
  - a. day 1, plate into T225: use 50mL of media.
  - b. day 2, change media, use 50mL of media
  - c. day 4, change media, use 100mL of media (if confluency is >50%)
  - d. day 6, change media, use 100mL of media (or harvest if ready)
  - e. day 7 or 8 (harvest when cells reach 6 x 10<sup>6</sup> cells/flask).

### D. Harvest

- 1) Pass cells 3-4 times until the desired cell number is achieved (primary cells will senesce after 4-5 passages).
- 2) Remove cells from flasks according to protocol described above under 'Sub-culture'.
- 3) Examine viability using Trypan blue staining (SOP TP-7).